

## Flow Cytometry

### Becton Dickinson LSR-II



3 laser (B/R/V), 2 scatter, 14 fluorochrome (6/3/6) LSR-II flow cytometer for Multiparameter analysis.

This model has a High Throughput System (HTS) for acquiring data from 96 well plates and FACSFlow Supply system for ease of use in operation.

Cytometer setup and tracking (CST) and FACSDiva 6.1.3 software are used to QC the instrument and acquire the data.

The standard configuration is found in the attached document: [Standard Filter Set](#)

For Training and Information on the LSRII contact: [elizabeth.king@lshtm.ac.uk](mailto:elizabeth.king@lshtm.ac.uk)

### Becton Dickinson FACSCalibur

The FACS Calibur is located in our Containment Level 3 laboratory for work with ACDP3 pathogens.

The 488nm (Ar) and 635nm red diode lasers allow 4 fluorescence and 2 light scatter parameter detection.

This model has a Carousel option, and the Macintosh Computer runs CellQuest v3.3 and Worklist Manager software for acquisition.

For Training and Information on the FACSCalibur contact the CL3 Laboratory Manager

## Flow Cytometric Data Analysis

There is a dedicated Macintosh Workstation located on the 6<sup>th</sup> floor specifically for the Analysis of Flow Cytometric data using FlowJo and Spice\*.

USB dongles for the use of FACSDiva and FCAP Array software on User's own computers are available for short-term loan.

A copy of Modfit LT is located on the Luminex PC workstation on the 6<sup>th</sup> floor.

## Magpix

There is a Magpix machine belonging to the Drakeley group that is available for use by Faculty Members, please contact [Elizabeth.king@lshtm.ac.uk](mailto:Elizabeth.king@lshtm.ac.uk) for further information.

## Further Information

A booking system is in operation to allow for equal opportunity of access and to aid cost recovery.

LSHTM Staff and Students can visit the following Network folder: J:/FACS Users.

Drop-in sessions are held between 9.00-9.30 every morning on the 6<sup>th</sup> floor during cytometer setup and tracking.

### Weblinks:

[Flow Cytometry a Basic Introduction - M.G. Ormerod](#)

[Current protocols in cytometry](#)

[Purdue University Cytometry Lab](#)

[Flow Cytometry UK](#)

[BD Spectra Viewer - requires Java](#)

[Coulter Spectra viewer - for download](#)

[eBioscience Spectra Viewer - requires Java](#)

[Invitrogen Spectra Viewer](#)

[Fluorish - panel designer](#)

[Novusbio](#) – panel designer

[FACSDiva Acquisition & Analysis software](#)

[FlowJo Analysis software](#)

[Modfit LT Analysis software](#)

\*[Spice Analysis software](#)

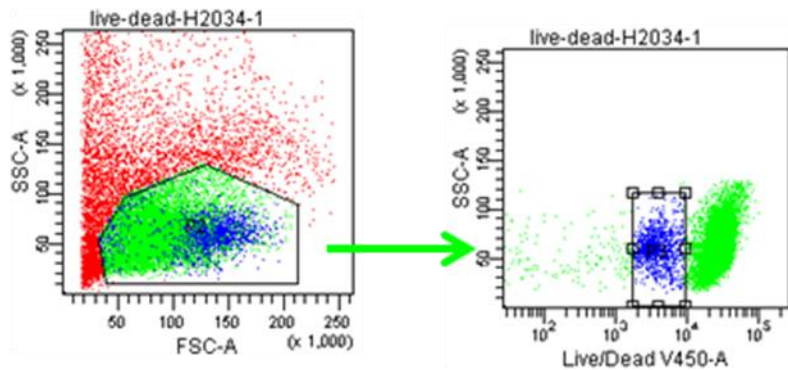
[CBA](#) – bead multiplexing

[Flow Cytomix - bead multiplexing](#)

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## Image Gallery

Live/Dead Staining of Cultured Human PBMCs analysed with FACS Diva:



Spleen from VertX (GFP IL-10 reporter) and Yeti (YFP IFN-g reporter) mice infected with *Plasmodium Yoelii* parasites analysed with FlowJo:

